Abstract: We describe cervids as potential reservoir hosts of *Babesia* EU1 and *B. divergens*. Both babesial parasites were found in roe deer. Sequence analysis of 18S rRNA showed 99.7% identity of roe deer *Babesia* EU1 with the human EU1 strain. *B. divergens* detected in cervids was 99.6% identical to bovine *B. divergens*. 

Keywords: 8SI/analysis/Babesia divergens/Babesia EU1/babesiae/Capreolus capreolus/deer/disease/host-parasite relationship/human/Malme/parasite/reservoir/roe deer/rRNA/sequence analysis
Cervids as Babesiae Hosts, Slovenia

Darja Duh,* Miroslav Petrovec,† and Tatjana Avsic-Zupanc*

We describe cervids as potential reservoir hosts of Babesia EU1 and B. divergens. Both babesial parasites were found in roe deer. Sequence analysis of 18S rRNA showed 99.7% identity of roe deer Babesia EU1 with the human EU1 strain. B. divergens detected in cervids was 99.6% identical to bovine B. divergens.

Human babesiosis is an emerging tick-transmitted disease caused by intraerythrocytic parasites of the genus Babesia. A bovine parasite, Babesia divergens has been implicated as the most common agent of this dangerous zoonosis in Europe (1). The life cycle of B. divergens is determined by cattle, the vertebrate host, and by European sheep ticks, Ixodes ricinus. Ticks are not only the vectors of B. divergens but also its most important non-bovine reservoir (2). Many questions regarding parasite epidemiology and biology and the host response to infection remain to be answered. Furthermore, molecular data for B. divergens are scarce; only 1 DNA sequence of this parasite from humans from mainland Europe has been recently deposited (3). Recently, 2 cases of human babesiosis have been reported in Italy and Austria. The etiologic agent was identified as Babesia EU1, a pathogen closely related to, but clearly distinct from, B. divergens (4). The distinction was based on analysis of the complete babesial 18S rRNA gene, which also showed that EU1 is most closely related to B. odocoilei, a parasite of white-tailed deer (Odocoileus virginianus) in the United States (5). I. ricinus, the most prevalent and widely distributed tick species in Europe, has already been implicated as the vector of EU1 (4,6). Moreover, I. ricinus has a wide range of vertebrate hosts and readily bites humans. Rapidly and accurately identifying the reservoir of Babesia EU1 will enable appraisal of the full range of disease control options.

The Study

We investigated 2 species of cervids shot by professional hunters from 1996 to 2000 in the vicinity of Ljubljana, Slovenia. DNA was extracted from spleen samples of 51 roe deer (Capreolus capreolus) and 30 red deer (Cervus elaphus), as previously described (6). Babesiae were detected in cervids by using specific nested polymerase chain reaction (PCR) that allowed discrimination between B. divergens and EU1. Primers were designed on the basis of alignment of complete 18S rRNA gene sequences of EU1, B. divergens, and B. odocoilei. With primers PIRO-A (7) and BABSr, a 600-bp babesial 18S rRNA gene was amplified with 5 µL of DNA and PCR Master Mix (Promega, Madison, WI, USA). One microliter of PCR product was used for nested PCR with either primer set PIRO-B/BOD and PIRO-B/BDV to detect 240 bp of 18S rRNA of EU1 and B. divergens, respectively. Both babesial parasites were detected in roe deer (76.5%); however, more animals were infected with B. divergens (54.9%) than Babesia EU1 (21.6%). Only 16.7% red deer were infected with B. divergens alone. Infection with babesial parasites did not differ significantly between sexes in either roe or red deer.

To assess DNA sequence homologies with EU1 from human and ticks, distinctive amplicons of the complete babesial 18S rRNA gene derived from cervids were cloned and sequenced. Parasite DNA from 2 roe deer that were found positive with a different set of nested primers was used in PCR with CRYPTO F and CRYPTO R (6). Amplicons were ligated into a plasmid vector (TOPO TA Cloning Kit for Sequencing, Invitrogen, Groningen, the Netherlands), and Escherichia coli–competent cells were transformed as instructed by the manufacturer. Plasmid DNA was purified from overnight cultures of selected colonies (Wizard Plus Minipreps DNA Purification System, Promega) and analyzed for inserts by restriction analysis with EcoR1 (Promega). Sequencing on both strands was carried out in an automated sequencer using BigDye Terminator Cycle Sequencing Ready Reaction Kit (PE Applied Biosystems, Foster City, CA, USA). Two clones were included in reactions with T3, T7, and internal primers to obtain complete gene sequence. All primers designed and used for this study are listed in the Table. Sequences were analyzed with computer programs of the Lasergene 1999 software package (DNASTAR, Madison, WI, USA) and submitted to GenBank to determine accession numbers. Homology search and alignment of the complete sequence of the babesial 18S rRNA gene from 1 roe deer showed 99.7% (5 nucleotide [nt] differences) identity with EU1 from a human patient and 99.8% (4 nt differences) identity with EU1 present in I. ricinus ticks from Slovenia. The complete sequence of the babesial 18S rRNA gene from another roe deer was, however, nearly identical (99.6%, 7 nt differences) to babesial parasite MO1 and B. divergens. Phylogenetic relationships of babesiae from roe deer and from other sources are shown in the Figure. By using TREECON software (8), a phylogenetic tree was constructed with the neighbor-joining
method, and topology of the tree was obtained with the K80 model. Support for the tree nodes was calculated with 1,000 bootstrap replicates.

Conclusions

*Babesia* EU1, a zoonotic pathogen, was the cause of human babesiosis recently reported by Herwaldt et al. (4). While *I. ricinus* was already implicated as a vector of EU1, no other information about biology, ecology, or geographic distribution of EU1 exists (4,6). Phylogenetic analysis based on comparing the complete 18S rRNA gene sequence of EU1 derived from humans and ticks with other babesial parasites showed that EU1 is more closely related to *B. odocoilei* than *B. divergens* (4). *B. odocoilei*, which is transmitted by *I. scapularis*, primarily infects white-tailed deer in the United States (5). Cases of fatal babesiosis were described in 2 other species of cervids, namely a zoo-housed caribou (*Rangifer tarandus caribou*) and an elk (*C. elaphus elaphus*) (9). Therefore, we tested 2 species of cervids from Slovenia as potential reservoir hosts of EU1.

By using specific nested PCR, the presence of EU1 was established in roe deer (21.6%) but not in red deer. In Slovenia, roe deer are widely distributed, and the population density is high. Their pasture comprises woodland, bushes, and even open meadows and fields (10). However, red deer were nearly extinct in Slovenia in the beginning of the 19th century. Although they were later imported from Austria, Poland, and Hungary, they are still less numerous and therefore harbor fewer ticks (10). The identity of babesial parasites from roe deer from Slovenia with EU1 was confirmed by cloning and sequencing the complete babesial 18S rRNA gene. The sequences obtained were 99.8% and 99.7% identical to the 18S rRNA genes of EU1 from ticks and humans, respectively. Since the habitat of roe deer is expanding in other European countries (10), additional studies are needed to determine whether roe deer are reservoir hosts of EU1 elsewhere in Europe.

Whereas the presence of EU1 in cervids was anticipated, detection of *B. divergens* in roe and red deer was surprising. With the exception of a single report of naturally acquired babesiosis caused by *B. divergens* in reindeer (*R. tarandus tarandus*), no data about cervids as reservoirs of *B. divergens* were available at the time of our research (11,12). Although *B. divergens* can infect cervids experimentally, animals experience only mild infections with low parasitemia (2,11). However, 54.9% of roe deer and 16.7% of red deer were infected with *B. divergens* in this study. Further cloning and sequencing of the complete 18S rRNA gene of the parasite indicated 99.6% (7 nt differences) identity with babesial parasite MO1 and *B. divergens*. MO1 was described as an etiologic agent of human babesiosis acquired in Missouri and was genetically almost identical to *B. divergens* (99.9% identity, 2 nt differences).

Figure. Phylogenetic relationships of representative babesiae deposited in GenBank and detected in this study, inferred from multiple sequence alignment of complete 18S rRNA gene. Accession numbers of babesiae: *Babesia* EU1 from *Ixodes ricinus* ticks, AY553915; babesiae from roe deer 1, AY572457; babesiae from roe deer 2, AY572456; *Babesia EU1* from human, AY046575; *B. divergens*, AY046576; *B. odocoilei*, AY046577; *Babesia MO1*, AY048113; *B. caballi*, Z15104; and *Theileria annulata*, M64243.

The number on each branch shows the percent occurrence in 1,000 bootstrap replicates.
differences), but the authors claimed that the parasites probably differ (13). However, piroplasms in abnormal hosts or hosts that are not generally considered primary hosts may have morphologic differences (12). In addition, high molecular identity of piroplasms does not necessarily mean that they have the same infectivity for different hosts.

A Babesia sp., tentatively called B. capreoli, was observed and described in red deer in Scotland (14) and sika deer (C. nippon) in Ireland (15). The parasite resembled B. divergens morphologically and antigenically. B. capreoli was suggested to be transmitted by I. ricinus ticks. The main difference between bovine B. divergens and these deer parasites is in their host specificity. Whereas B. divergens can infect a wide range of animals after splenectomy, including some deer species, various nonhuman primates, gerbils, and humans, B. capreoli can apparently infect only cervids and perhaps sheep (15). Nevertheless, with no deposited sequence of 18S rRNA of B. capreoli, the identity of B. divergens from roe and red deer from Slovenia is uncertain.

The finding that roe and red deer may be reservoirs for B. divergens has serious implications. Future research must determine if parasites from cervids share biologic characteristics with B. divergens, such as infectivity to cattle and humans and transmission by I. ricinus.

Acknowledgments

We thank Jeremy S. Gray for critical review and helpful comments on the manuscript.

Ms. Duh is a PhD student at the Medical Faculty, Ljubljana, Slovenia. She is currently working at the Institute of Microbiology and Immunology. Her research is primarily based on babesiae and other tickborne pathogens.

References


Address for correspondence: Darja Duh, Institute of Microbiology and Immunology, Medical Faculty, Zaloška 4, 1000 Ljubljana, Slovenia; fax: 386-1-543-7401; email: darja.duh@mf.uni-lj.si

www.cdc.gov/eid

To receive tables of contents of new issues send an email to listserv@cdc.gov with subscribe eid-toe in the body of your message.